

Relief from a heavy heart: redox-sensitive NF- κ B as a therapeutic target in managing cardiac hypertrophy

Chandan K. Sen and Sashwati Roy

Laboratory of Molecular Medicine, Dorothy M. Davis Heart and Lung Research Institute,
Department of Surgery, The Ohio State University Medical Center, Columbus, Ohio

CARDIAC HYPERTROPHY is both an adaptive response to chronic pressure overload and a key risk factor in patients with hypertension (6). On one hand, cardiac hypertrophy may normalize wall tension, whereas on the other it may progress to a decompensated state causing overt heart failure, a common end stage of numerous cardiac disorders. Mechanisms of progression to heart failure include renal sodium and water retention, neurohumoral activation, and changes in cell signaling in the myocardial tissue (5, 18).

Fifteen years ago, Sen and associates (36) reported that mechanisms involved in the development or the regression of myocardial hypertrophy cannot be fully explained as responses to blood pressure control alone. They postulated that the development of hypertrophy is initiated by a mechanical or humoral signal to the myocardium, which in turn produces a soluble factor that triggers protein synthesis and initiates myocardial growth. Myotrophin was identified as a novel protein that was present in human, dog, and rat hypertrophied hearts. In neonatal cardiac myocytes, myotrophin increased cell surface area and caused myofibrils organization. Thus myotrophin emerged as a putative causative factor supporting cardiac hypertrophy (36). Later, Sen and associates (37) went on to identify myotrophin as a unique rel/nuclear factor (NF)- κ B interacting protein in the rat heart. One of the ankyrin repeats of myotrophin was highly homologous specifically to those of I κ B α /rel ankyrin repeats. Putative consensus phosphorylation sites for protein kinase C and casein kinase II, which were observed in I κ B α proteins, were identified in myotrophin (37). It is now known that myotrophin stimulates protein synthesis and myocardial cell growth associated with increased levels of hypertrophy marker genes. A basal level of myotrophin exists in both the cytoplasm as well as the nucleus under normal conditions. In response to cyclic stretch, however, myotrophin levels elevate in the nucleus. Exposure of excised beating hearts to high pressure induces myotrophin gene expression. Soon myotrophin- κ B DNA interaction emerged as an important step in initiating cardiac hypertrophy (11). Concurrent report by the same group demonstrating that the protein kinase C-*IKK*-NF κ B pathway is involved in mediating myotrophin-induced hypertrophic response in cardiomyocytes represents a landmark progress identifying NF- κ B as a molecular checkpoint regulating cardiac hypertrophy. NF- κ B-dependent induction of the hypertrophic genes atrial natriuretic factor (ANF) and *c-myc* in cardiomyocytes marks a key mechanism underlying the hypertrophic effects of myotrophin (10).

As the evidence in support of the myotrophin-NF- κ B paradigm mounted based on the study of rodent models, a clear

need to develop the significance in a clinical heart failure setting arose. Plasma myotrophin levels were measured in 120 patients with heart failure and 130 age- and gender-matched normal controls (23). Whereas the results do not establish a causal relationship between myotrophin and heart failure, the observation reporting early activation of the myotrophin system in heart failure represents a significant confidence-building piece for the myotrophin-NF- κ B paradigm in cardiac hypertrophy (23). Overexpression of heart-specific myotrophin in transgenic mice causes cardiac hypertrophy that progress to heart failure, similar to changes in human heart failure. Such hypertrophy is associated with increased expression of proto oncogenes, hypertrophy marker genes, growth factors, and cytokines, with symptoms that functionally and morphologically mimic those of human cardiomyopathy (27, 28).

The first direct evidence suggesting that NF- κ B activation is required for the development of cardiac hypertrophy in vivo was obtained only recently. In an experimental model of aortic banding-induced cardiac hypertrophy, genetic and antioxidant strategies to arrest NF- κ B activity attenuated banding-induced increase in heart-to-body weight ratio (17). Compared with pharmacological approaches, genetic tools to manipulate any given signaling pathway are more specific and thus more valuable in the laboratory. Once the hypothesis is tested and the principles unveiled, taking experimental findings to the bedside rely mostly on pharmacological approaches. Pharmacologically, inducible NF- κ B activation may be repressed by strategies antagonizing oxidants (e.g., antioxidants), inhibiting I κ B phosphorylation and degradation (e.g., sodium salicylate and acetylsalicylic acid or aspirin), or preventing binding of nuclear NF- κ B protein to the κ B site (e.g., aurine tricarboxylic acid). The antioxidant approach is effective in vivo (19, 20). In this issue of the *American Journal of Physiology-Heart and Circulatory Physiology*, Sen and associates demonstrate that the antioxidant pyrrolidine dithiocarbamate (PDTC) inhibits NF- κ B activation in vivo and regressed cardiac hypertrophy in spontaneously hypertensive rats. The effect of PDTC was dependent on NF- κ B and independent of hypertension (12). These findings underscore the potential of redox-active inhibitors of inducible NF- κ B activity as therapeutic candidates in the management of cardiac hypertrophy.

During the last decade, as compelling evidence continued to accumulate supporting that reactive oxygen species (ROS) serve as cellular messenger molecules (3, 4, 26, 32), NF- κ B has emerged as one of the most well-studied molecular checkpoints, the inducible activation of which may be reliably inhibited by redox-active agents (8, 14, 16, 29, 32, 34, 35). PDTC became a common experimental tool to inhibit inducible NF- κ B activation. First synthesized in the mid-1800s, dithiocarbamates have found applications in the pharmaceutical industries because of their metal binding and antioxidant prop-

Address for reprint requests and other correspondence: C. K. Sen, 512 Davis Heart and Lung Research Institute, The Ohio State Univ. Medical Center, 473 W. 12th Ave., Columbus, OH 43210 (E-mail: sen-1@medctr.osu.edu).

erties (38, 39). Despite promising outcomes in the treatment of autoimmune deficiency syndrome (25) and cancer (7), dithiocarbamates do not seem to have found their way to clinical testing for more than a decade. Rationale for this cautious approach perhaps remain embedded in the historical background of dithiocarbamates as agricultural insecticides and fungicides (13). Dithiocarbamates, including PDTC, may undergo copper-catalyzed oxidation to form the corresponding thiuram disulfides, which are cytotoxic (2).

ROS function as second messengers signaling for the hypertrophy of cardiac myocytes (1, 43). TNF- α -induced myocyte hypertrophy is mediated through NF- κ B activation via the generation of ROS (15). These observations rationally lead to the hypothesis that antioxidant-based therapies, dually targeting ROS and NF- κ B, may help prevent cardiac hypertrophy. Among the thiol-based antioxidants that have proven ability to inhibit inducible NF- κ B activity (29, 31–33), *N*-acetylcysteine (NAC) and α -lipoic acid (ALA) have a sound track record for safe and effective clinical use. Biochemical studies predict that based on the mechanisms of thiol regeneration, ALA is a more effective thiol antioxidant than NAC (24, 30). ALA defends against certain risk factors of cardiovascular disease (42). However, the insulin mimetic properties of ALA have made it a more suited candidate for the treatment of diabetes (24). Further studies are necessary to establish the benefits of ALA to patients suffering from cardiac disorders.

Oral and intravenous use of NAC has an extensive history of clinical studies addressing a wide range of disorders. In patients with unstable angina pectoris and a threat of infarct, the intravenous or oral administration of NAC in association with nitroglycerin effectively decreases the risk of worsening, mainly by preventing the occurrence of acute myocardial infarction (21). As an effective antioxidant in a clinical setting, NAC attenuates myocardial oxidative stress in the hearts of patients subjected to cardiopulmonary bypass and cardioplegic arrest (40). NAC improved bradycardic and tachycardic baroreflex responses in spontaneously hypertensive rats without modifying catecholamine responses (9). In addition to these favorable effects on the cardiovascular system, NAC has been tested in the laboratory for its effect on cardiac hypertrophy. NAC prevents cardiomyocytes hypertrophy in vitro (41). Angiotensin II-induced ANF expression in neonatal rat cardiac myocytes was also inhibited by NAC. Furthermore, NAC inhibited angiotensin II-induced cardiac hypertrophy in vivo (22). Taken together, the current state of knowledge warrants clinical testing of antioxidant therapies targeting inhibition of inducible NF- κ B activation in the heart for the treatment of cardiac hypertrophy and related heart failure.

GRANTS

Research on tissue injury and repair is supported by National Institutes of Health Grants RO1 HL-73087, NS-42617, and GM-69589 (to C. K. Sen).

REFERENCES

- Aikawa R, Nagai T, Tanaka M, Zou Y, Ishihara T, Takano H, Hasegawa H, Akazawa H, Mizukami M, Nagai R, and Komuro I. Reactive oxygen species in mechanical stress-induced cardiac hypertrophy. *Biochem Biophys Res Commun* 289: 901–907, 2001.
- Burkitt MJ, Bishop HS, Milne L, Tsang SY, Provan GJ, Nobel CS, Orrenius S, and Slater AF. Dithiocarbamate toxicity toward thymocytes involves their copper-catalyzed conversion to thiuram disulfides, which oxidize glutathione in a redox cycle without the release of reactive oxygen species. *Arch Biochem Biophys* 353: 73–84, 1998.
- Demple B. Redox signaling and gene control in the *Escherichia coli* SoxRS oxidative stress regulon—a review. *Gene* 179: 53–57, 1996.
- Finkel T. Redox-dependent signal transduction. *FEBS Lett* 476: 52–54, 2000.
- Frey N and Olson EN. Cardiac hypertrophy: the good, the bad, and the ugly. *Annu Rev Physiol* 65: 45–79, 2003.
- Frohlich ED. Cardiac hypertrophy in hypertension. *N Engl J Med* 317: 831–833, 1987.
- Gandara DR, Perez EA, Weibe V, and De Gregorio MW. Cisplatin chemoprotection and rescue: pharmacologic modulation of toxicity. *Semin Oncol* 18: 49–55, 1991.
- Ginn-Pease ME and Whisler RL. Redox signals and NF- κ B activation in T cells. *Free Rad Biol Med* 25: 346–361, 1998.
- Girouard H, Denault C, Chulak C, and de Champlain J. Treatment by *N*-acetylcysteine and melatonin increases cardiac baroreflex and improves antioxidant reserve. *Am J Hypertens* 17: 947–954, 2004.
- Gupta S, Purcell NH, Lin A, and Sen S. Activation of nuclear factor- κ B is necessary for myotrophin-induced cardiac hypertrophy. *J Cell Biol* 159: 1019–1028, 2002.
- Gupta S and Sen S. Myotrophin- κ B DNA interaction in the initiation process of cardiac hypertrophy. *Biochim Biophys Acta* 1589: 247–260, 2002.
- Gupta S, Young D, and Sen S. Inhibition of NF- κ B induces regression of cardiac hypertrophy, independent of blood pressure control, in spontaneously hypertensive rats. *Am J Physiol Heart Circ Physiol* 288: H20–H29, 2005. First published March 4, 2005; 10.1152/ajpheart.00082.2005.
- Hayes WJ. *Pesticides Studied in Man*. Baltimore, MD: Williams and Wilkins, 1982.
- Higuchi M, Manna SK, Sasaki R, and Aggarwal BB. Regulation of the activation of nuclear factor κ B by mitochondrial respiratory function: evidence for the reactive oxygen species-dependent and -independent pathways. *Antiox & Redox Signal* 4: 945–955, 2002.
- Higuchi Y, Otsu K, Nishida K, Hirotsu S, Nakayama H, Yamaguchi O, Matsumura Y, Ueno H, Tada M, and Hori M. Involvement of reactive oxygen species-mediated NF- κ B activation in TNF- α -induced cardiomyocyte hypertrophy. *J Mol Cell Cardiol* 34: 233–240, 2002.
- Janssen-Heininger YM, Poynter ME, and Baeuerle PA. Recent advances towards understanding redox mechanisms in the activation of nuclear factor κ B. *Free Radic Biol Med* 28: 1317–1327, 2000.
- Li Y, Ha T, Gao X, Kelley J, Williams DL, Browder IW, Kao RL, and Li C. NF- κ B activation is required for the development of cardiac hypertrophy in vivo. *Am J Physiol Heart Circ Physiol* 287: H1712–H1720, 2004.
- Lips DJ, deWindt LJ, van Kraaij DJ, and Doevendans PA. Molecular determinants of myocardial hypertrophy and failure: alternative pathways for beneficial and maladaptive hypertrophy. *Eur Heart J* 24: 883–896, 2003.
- Liu SF, Ye X, and Malik AB. Inhibition of NF- κ B activation by pyrrolidine dithiocarbamate prevents in vivo expression of proinflammatory genes. *Circulation* 100: 1330–1337, 1999.
- Liu SF, Ye X, and Malik AB. Pyrrolidine dithiocarbamate prevents I- κ B degradation and reduces microvascular injury induced by lipopolysaccharide in multiple organs. *Mol Pharmacol* 55: 658–667, 1999.
- Marchetti G, Lodola E, Licciardello L, and Colombo A. Use of *N*-acetylcysteine in the management of coronary artery diseases. *Cardiology* 44: 633–637, 1999.
- Nakagami H, Takemoto M, and Liao JK. NADPH oxidase-derived superoxide anion mediates angiotensin II-induced cardiac hypertrophy. *J Mol Cell Cardiol* 35: 851–859, 2003.
- O'Brien RJ, Loke I, Davies JE, Squire IB, and Ng LL. Myotrophin in human heart failure. *J Am Coll Cardiol* 42: 719–725, 2003.
- Packer L, Roy S, and Sen CK. Alpha-lipoic acid: a metabolic antioxidant and potential redox modulator of transcription. *Adv Pharmacol* 38: 79–101, 1997.
- Reisinger EC, Kern P, Ernst M, Bock P, Flad HD, and Dietrich M. Inhibition of HIV progression by dithiocarb. *Germ DTC Study Group Lancet* 335: 679–682, 1990.
- Rhee SG. Redox signaling: hydrogen peroxide as intracellular messenger. *Exp Molec Med* 31: 53–59, 1999.
- Sarkar S, Chawla-Sarkar M, Young D, Nishiyama K, Rayborn ME, Hollyfield JG, and Sen S. Myocardial cell death and regeneration during progression of cardiac hypertrophy to heart failure. *J Biol Chem* 279: 52630–52642, 2004.

28. Sarkar S, Leaman DW, Gupta S, Sil P, Young D, Morehead A, Mukherjee D, Ratliff N, Sun Y, Rayborn M, Hollyfield J, and Sen S. Cardiac overexpression of myotrophin triggers myocardial hypertrophy and heart failure in transgenic mice. *J Biol Chem* 279: 20422–20434, 2004.
29. Sen CK. Cellular thiols and redox-regulated signal transduction. *Curr Top Cell Regul* 36: 1–30, 2000.
30. Sen CK. Nutritional biochemistry of cellular glutathione. *J Nutr Biochem* 8: 660–672, 1997.
31. Sen CK. Redox signaling and the emerging therapeutic potential of thiol antioxidants. *Biochem Pharmacol* 55: 1747–1758, 1998.
32. Sen CK and Packer L. Antioxidant and redox regulation of gene transcription. *FASEB J* 10: 709–720, 1996.
33. Sen CK and Packer L. Thiol homeostasis and supplements in physical exercise. *Am J Clin Nutr* 72: 653S–669S, 2000.
34. Sen CK, Roy S, and Packer L. Involvement of intracellular Ca²⁺ in oxidant-induced NF-kappa B activation. *FEBS Lett* 385: 58–62, 1996.
35. Sen CK, Sies H, and Baeuerle PA. Antioxidant and Redox Regulation of Genes. San Diego, CA: Academic, 2000, p. 556.
36. Sen S, Kundu G, Mekhail N, Castel J, Misono K, and Healy B. Myotrophin: purification of a novel peptide from spontaneously hypertensive rat heart that influences myocardial growth. *J Biol Chem* 265: 16635–16643, 1990.
37. Sivasubramanian N, Adhikary G, Sil PC, and Sen S. Cardiac myotrophin exhibits rel/NF-kappa B interacting activity in vitro. *J Biol Chem* 271: 2812–2816, 1996.
38. Somers PK, Medford RM, and Saxena U. Dithiocarbamates: effects on lipid hydroperoxides and vascular inflammatory gene expression. *Free Radic Biol Med* 28: 1532–1537, 2000.
39. Thorn GD and Ludwig RA. *The Dithiocarbamates and Related Compounds*. Amsterdam: Elsevier, 1962.
40. Tossios P, Bloch W, Huebner A, Raji MR, Dodos F, Klass O, Suedkamp M, Kasper SM, Hellmich M, and Mehlhorn U. N-acetylcysteine prevents reactive oxygen species-mediated myocardial stress in patients undergoing cardiac surgery: results of a randomized, double-blind, placebo-controlled clinical trial. *J Thorac Cardiovasc Surg* 126: 1513–1520, 2003.
41. Tu VC, Bahl JJ, and Chen QM. Signals of oxidant-induced cardiomyocyte hypertrophy: key activation of p70 S6 kinase-1 and phosphoinositide 3-kinase. *J Pharmacol Exp Ther* 300: 1101–1110, 2002.
42. Wollin SD and Jones PJ. Alpha-lipoic acid and cardiovascular disease. *J Nutr* 133: 3327–3330, 2003.
43. Xie Z, Kometiani P, Liu J, Li J, Shapiro JJ, and Askari A. Intracellular reactive oxygen species mediate the linkage of Na⁺/K⁺-ATPase to hypertrophy and its marker genes in cardiac myocytes. *J Biol Chem* 274: 19323–19328, 1999.

