

Topical Lyophilized Targeted Lipid Nanoparticles in the Restoration of Skin Barrier Function following Burn Wound

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Lyophilized keratinocyte-targeted nanocarriers (TLN_k) loaded with locked nucleic acid (LNA) modified anti-miR were developed for topical application to full thickness burn injury. TLN_k were designed to selectively deliver LNA-anti-miR-107 to keratinocytes using the peptide sequence ASKAIQVFLLAG. TLN_k employed DOTAP/DODAP combination pH-responsive lipid components to improve endosomal escape. To minimize interference of clearance by non-targeted cells, especially immune cells in the acute wound microenvironment, surface charge was neutralized. Lyophilization was performed to extend the shelf life of the lipid nanoparticles (LNPs). Encapsulation efficiency of anti-miR in lyophilized TLN_k was estimated to be 96.54%. Cargo stability of lyophilized TLN_k was tested. After 9 days of loading with anti-miR-210, TLN_k was effective in lowering abundance of the hypoxamiR miR-210 in keratinocytes challenged with hypoxia. Keratinocyte uptake of DiD-labeled TLN_k was selective and exceeded 90% within 4 hr. Topical application of hydrogel-dispersed lyophilized TLN_k encapsulating LNA anti-miR-107 twice a week significantly accelerated wound closure and restoration of skin barrier function. TLN_{k/anti-miR-107} application depleted miR-107 and upregulated dicer expression, which accelerated differentiation of keratinocytes. Expression of junctional proteins such as claudin-1, loricrin, filaggrin, ZO-1, and ZO-2 were significantly upregulated following TLN_{k/anti-miR-107} treatment. These LNPs are promising as topical therapeutic agents in the management of burn injury.

INTRODUCTION

Lipid nanoparticles (LNPs) are promising as carriers of nucleic acid therapeutics. They have been used in cancer treatment for delivering targeted agents,¹ antisense oligonucleotides,^{2,3} small interfering RNA (siRNA),⁴⁻⁷ mRNA,⁸ and DNA inhibitors.⁹ Particle size and surface modifications are important parameters for LNPs.^{10,11} Cell targeting can be achieved by conjugating antibodies to the LNP surface.¹¹ Although such approach is effective *in vitro*, *in vivo* applications are fraught with the risk of immunogenicity following repeated injec-

tions.¹² Furthermore, barriers such as high cost, regulatory hurdles, and limited shelf life complicate the translational path.¹¹ Nonetheless, both therapeutic as well as diagnostic nanoparticles are currently undergoing clinical testing.¹³

Skin is a promising route for drug delivery offering the option to evade the first-pass effect of the liver that can prematurely metabolize drugs.¹⁴ Small LNPs (<10 nm) may directly penetrate the stratum corneum of viable human skin.¹⁵ Meanwhile, larger LNPs (10–200 nm) access the skin via hair follicle openings, especially after massage.¹⁵ These larger particles traverse the epidermis through shunt pathways generated by sweat glands and hair shafts. Such a first-generation delivery system is effective in delivering lipophilic small molecules.¹⁴ The low density of hair follicles (32 follicles/cm²) relative to the whole skin surface represents a limitation of the delivery system.¹⁶ Current delivery systems rely on chemical enhancers, non-cavitational ultrasound and iontophoresis, as well as additional measures such as microneedles or thermal abrasion to penetrate the stratum corneum.¹⁴ Targeted delivery of therapeutic cargo across intact stratum corneum remains a challenge.

In wounds, although skin barrier function is breached, abundance of inflammatory cells at the site of injury poses a different set of challenges. Phagocytic clearance of nanoparticles poses an issue for delivery.¹⁷⁻¹⁹ In this work, we sought to develop a novel delivery platform based on lyophilized keratinocyte targeting LNPs (TLN_k) to facilitate keratinocyte-specific delivery of oligonucleotides such as locked nucleic acid (LNA)-modified anti-microRNAs (miRNAs). Hydrogel-assisted delivery of lyophilized TLN_k was shown to be robust, offering translational advantages and a longer shelf life in wound care.

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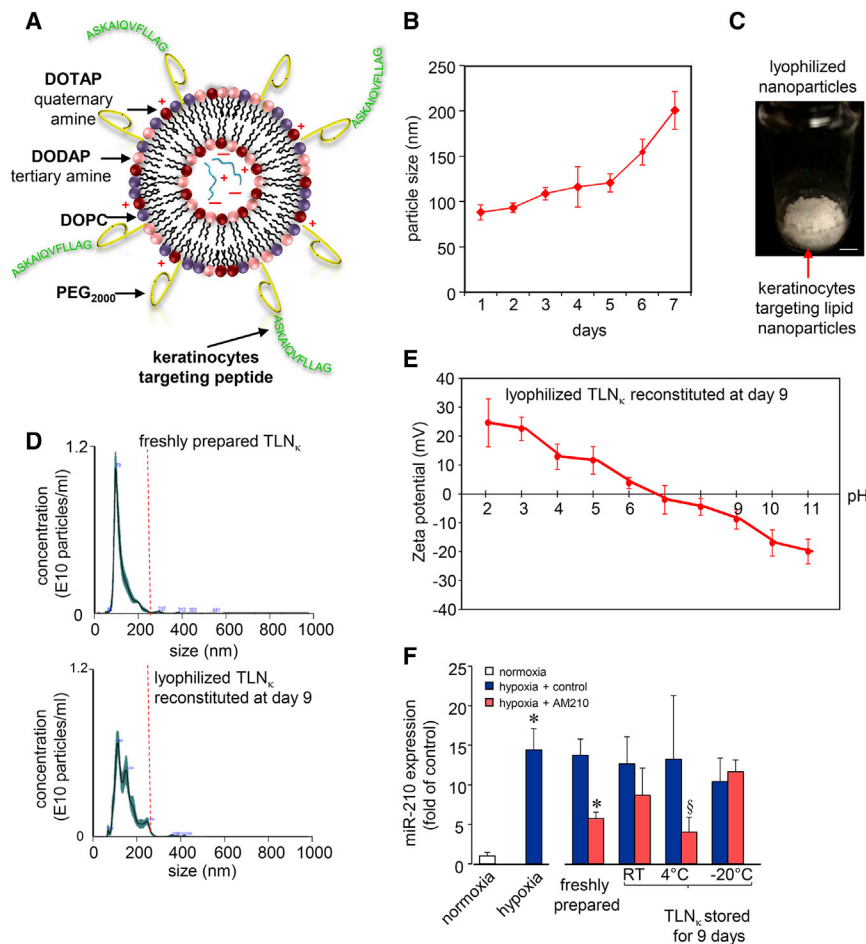


Figure 1. Characterization of the Keratinocytes Targeting Lyophilized Lipid Nanoparticles

(A) Schematic representation of keratinocytes targeting lipid nanoparticles (TLN_κ). (B) Change of TLN_κ size due to aqueous storage condition at 4°C over a period of 7 days. (C) Representative photograph of the lyophilized keratinocytes targeting lipid nanoparticles. (D) Representative nanoparticle tracking analysis (NanoSight) showing particle size and concentration of freshly prepared TLN_κ and reconstituted lyophilized TLN_κ after 9 days of storage under 4°C (n = 4). (E) The zeta potential of the TLN_κ at different pH. (F) *miR-210* expression in HaCaT cells exposed to normoxia and hypoxia 24 hr after delivery of TLN_κ/anti-*miR-210*. Data expressed as mean ± SD (n = 4). §p < 0.05, *p < 0.001 compared to hypoxia, ANOVA.

hypoxia, showed elevated *miR-210* expression. TLN_κ loaded with anti-*miR-210*, stored at 4°C for 9 days, significantly inhibited *miR-210* response in cells exposed to hypoxia. Storage of LNPs at 4°C yielded the best result, compared to storage at room temperature or at -20°C (Figure 1F).

In Vitro Targeting Efficiency of TLN_κ

To test specificity of targeted cell delivery of TLN_κ, *in vitro* experiments were carried out using different cell lines, including keratinocytes (HaCaT cells), endothelial cells (HMEC), fibroblasts (BJ), and differentiated monocytes (THP-1). 1,1-Dioctadecyl-3,3,3,3-tetramethylindodicarbocyanine (DiD), a lipophilic

fluorescent dye, was incorporated into the LNPs as described in the [Materials and Methods](#).²³ Cells were exposed to DiD-labeled TLN_κ for 4 hr followed by imaging and flow cytometric analyses. The uptake of DiD-labeled TLN_κ was significantly higher in keratinocytes compared to that of non-targeted LNPs (nTLN_κ without the targeting peptide) that served as control (Figure 2A). Flow cytometric data showed more than 90% keratinocyte uptake of the DiD-labeled TLN_κ, unlike that in endothelial cells (HMEC) or in fibroblasts (BJ) (Figure 2B). Differentiated monocytes (THP-1) appeared promiscuous, demonstrating 14% uptake of even nTLN_κ. Nonetheless, delivery of TLN_κ to keratinocytes was over 4-fold higher compared to uptake by monocytic cells.

In Vivo Targeting Efficiency of TLN_κ

To evaluate targeting efficiency of TLN_κ *in vivo*, full thickness burn wound was developed on the dorsal skin of mice. Full thickness burn wound was confirmed by H&E staining (Figure S1A). Lyophilized DiD-labeled TLN_κ was applied with hydrogel on the burn wound (Figure 3A). Lyophilized TLN_κ was readily dispersed in hydrogel within 30 s. The burn wound tissue was harvested after 24 hr of injury. To assess anti-*miR* delivery efficiency of TLN_κ *in vivo*,

RESULTS

Preparation and Characterization of Lyophilized TLN_κ

TLN_κ was prepared using a keratinocyte-targeting peptide sequence ASKAIQVFLLAG (A5G33).²⁰ The peptide was conjugated to 2-dis-tearoyl-*sn*-glycero-3-phosphoethanolamine-N-[amino(polyethylene glycol)-2000] (DSPE-PEG₂₀₀₀-Amine) and incorporated onto the surface of LNPs (Figure 1A). Colloidal stability of the LNPs was studied under storage condition. Over a period of 6 days, the particle size doubled from 88 nm to >200 nm (Figure 1B). To address such limitation, TLN_κ was lyophilized with addition of a lyoprotectant as described in the [Materials and Methods](#) (Figure 1C). The lyophilized keratinocyte-targeted LNPs when stored at 4°C retained their size when reconstituted after 9 days (Figure 1D). Zeta potential of these reconstituted TLN_κ ranged from +10 mV to -10 mV in the pH range of 6–8 (Figure 1E). Encapsulation of anti-*miRNA* in lyophilized TLN_κ after 9 days at 4°C was found to be 96.54%. In order to assess cargo stability of lyophilized TLN_κ, *in vitro* transfection was performed with TLN_κ loaded with anti-*miR-210*. Anti-*miR-210* was chosen as test cargo based on our previous publications demonstrating that in ischemic wounds, elevated *miR-210* compromises wound healing.^{21,22} Human immortalized keratinocytes cells, when subjected to

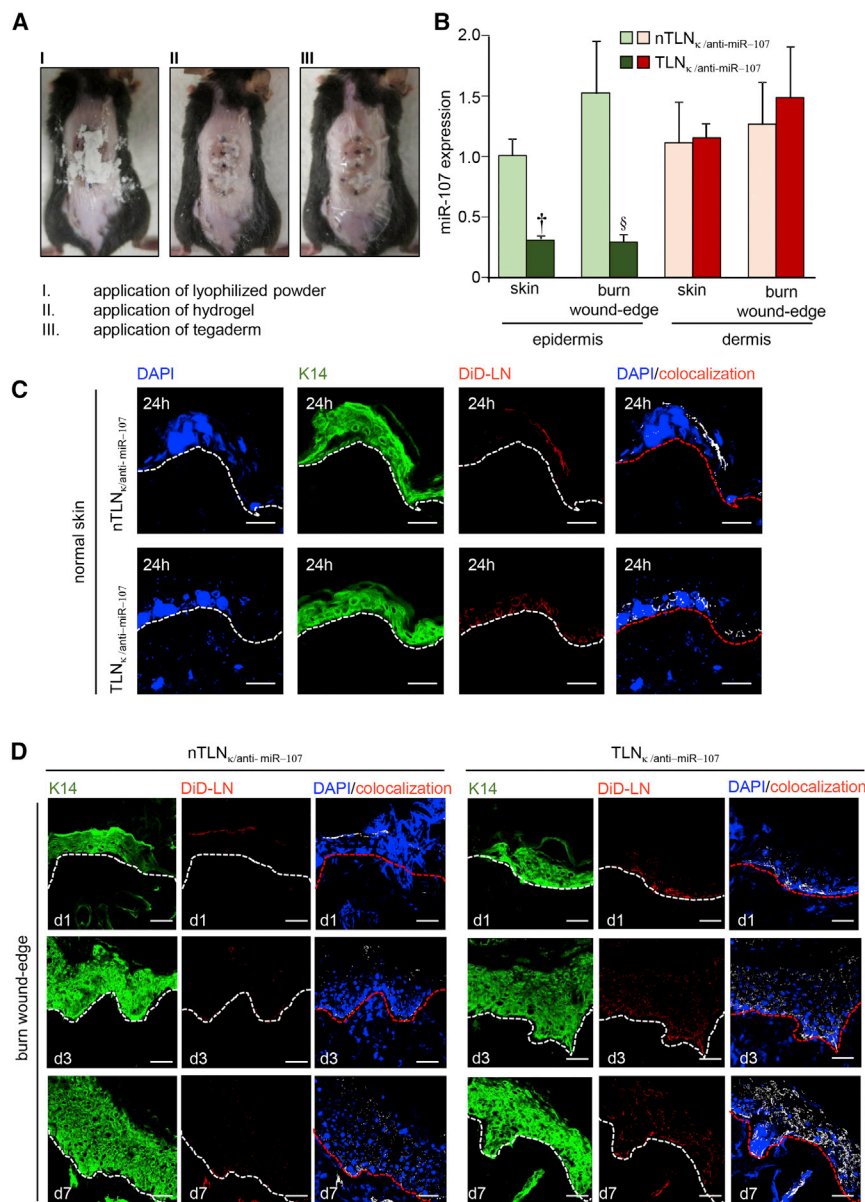


Figure 3. In Vivo Targeting Efficiency of TLN_κ

(A) Digital photomicrographs showing (I) application of lyophilized TLN_κ powder on the dorsal skin of mice post-burn; (II) application of 3M hydrogel to cover and reconstitute the dry powder; (III) application of 3M Tegaderm to cover and strap the dressing from falling off. (B) *miR-107* expression from laser microdissected epidermis of murine skin and wound-edge tissue 24 hr after application of lyophilized TLN_κ containing anti-*miR-107*. Data expressed as mean ± SEM (n = 3), §p < 0.05, †p < 0.01, ANOVA compared to nTLN_κ/anti-*miR-107*. (C) Confocal microscopic images showing localization of the DiD-labeled nanoparticles (red) in the epidermis 24 hr after application of nTLN_κ/anti-*miR-107* and TLN_κ/anti-*miR-107*. The sections were counterstained with K14 (green) and DAPI (blue). (D) Confocal microscopic images showing colocalization (white dots) of the DiD-labeled nanoparticles (red) in the burn wound-edge epidermis stained with K14 (green) at days 1, 3, and 7. The sections were counterstained with DAPI (blue). nTLN_κ/anti-*miR-107* and TLN_κ/anti-*miR-107* were applied as mentioned in the Materials and Methods. Scale bars, 50 μm.

first report on cell-targeting LNPs in a non-cancer health care application.³² According to a recent report from The Center for Drug Evaluation and Research (CDER) within the US Food and Drug Administration (FDA), 234 investigational new drug (IND) applications containing nanomaterials have been submitted, and 34 new drug applications (NDAs) have been approved by the office.³³ Liposomes account for 61% of nanomaterials targeting cancer therapeutics having a size less than 300 nm.³³ The following targeted nanoparticles are in clinical trial: MCC-465, MBP-426, SGT-53, SGT-94, BIND-014, MM-302, TargomiRs, CALAA-01, Cornell Dot, and ND-L02 s0201.^{34,35} Nanoparticles are frequently targeted to cancer cells either through the transferrin receptor or folate receptor.^{36–38} In addition, carbohydrates such as lacto bionic acid have been used to target hepatic tumor cells by binding to their asialoglycoprotein receptors.³⁹

Epidermal Junctional Proteins

Junctional proteins help establish the barrier function of the repaired skin. Topical application of lyophilized TLN_κ/anti-*miR-107* significantly upregulated the expression of claudin-1, loricrin, filaggrin, ZO-1, and ZO-2 (Figure 5) compared to lyophilized TLN_κ/scramble or lyophilized nTLN_κ/anti-*miR-107*.

DISCUSSION

Lipid nanomaterials with specific biological functions are of therapeutic value. In this work, we report a novel LNP formulation of anti-*miR* that when applied to the skin topically may penetrate the stratum corneum to specifically target keratinocytes for cargo delivery. The LNPs can be lyophilized to extend the shelf life. This work constitutes the

Polysaccharides, such as hyaluronic acid or chitosan, have shown promising results in targeting tumor extracellular environment.⁴⁰ However, the cellular uptake and delivery efficiency are difficult to predict as they depend on the payload-related variables.⁴⁰ Mannose and galactose derivatives are often used for targeting macrophages through the C-type lectin receptors, which are expressed by all the cells of myeloid origin.^{41,42} The targeting ability of functionalized nanoparticles may be lost in a biological environment.⁴³ Even when it does work, efficiency of targeting is highly limited and was estimated to be 30%.⁴⁴ Some carbohydrate nanocarriers (mono/oligosaccharides and/or polysaccharides) used in biomedical application are reported to reduce unspecific protein adsorption and increases the circulation

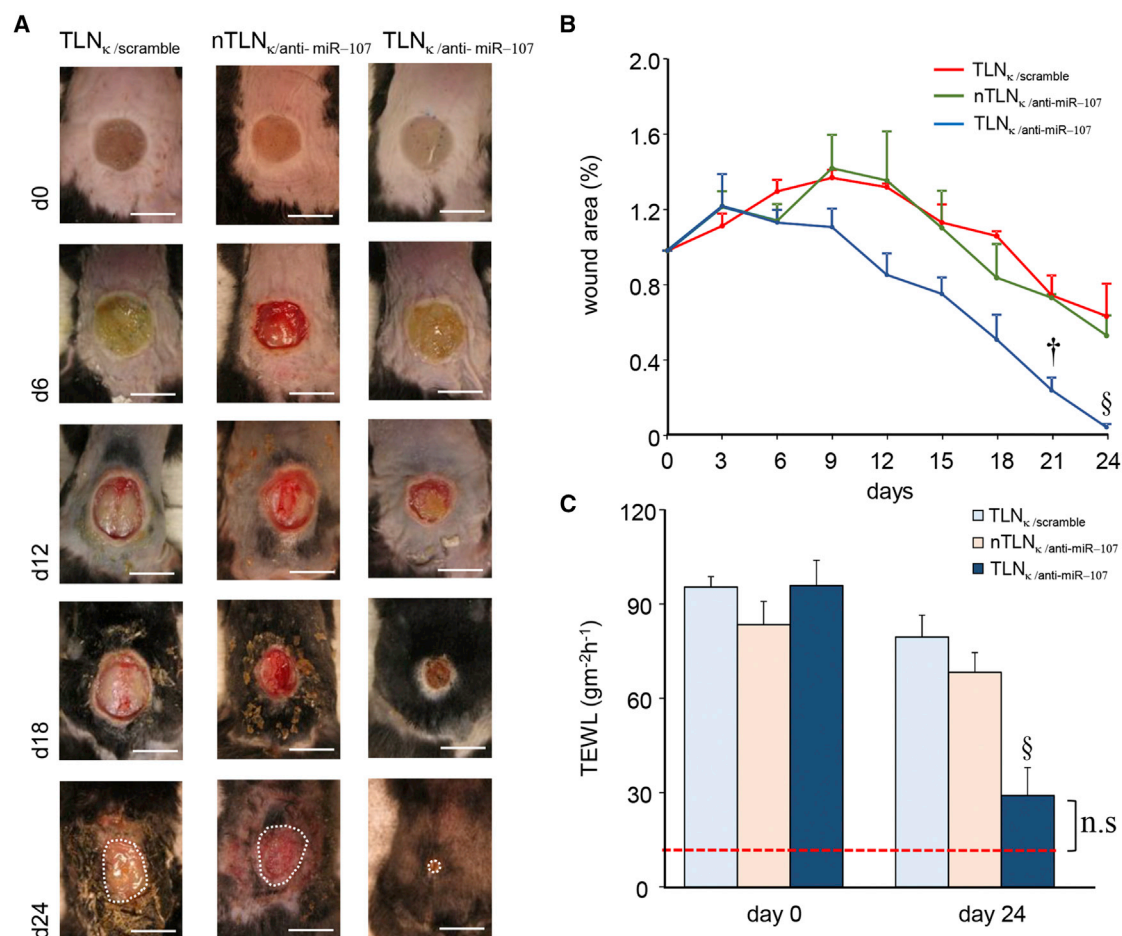


Figure 4. TLN_κ/anti-miR-107 Accelerates Wound Closure and Facilitates in Re-establishing Skin Barrier Function

(A) Digital photographs of the full thickness burn wound at day 0, 6, 12, 18, and 24 days after topical application of TLN_κ/scramble, nTLN_κ/anti-miR-107, and TLN_κ/anti-miR-107. The white dashed lines indicate the wound area. Scale bars, 1 cm. (B) Wound closure after topical application of TLN_κ/scramble, nTLN_κ/anti-miR-107, and TLN_κ/anti-miR-107 was quantified by digital planimetry. Data expressed as mean \pm SEM (n = 4), $\$p < 0.05$, $\dagger p < 0.01$ compared to day 0 (d0), ANOVA. (C) Transepidermal water loss at day 0 and at day 24 after delivery of TLN_κ/scramble, nTLN_κ/anti-miR-107, and TLN_κ/anti-miR-107 was plotted graphically. Data expressed as mean \pm SEM (n = 4), $\$p < 0.05$ compared to d0, ANOVA.

time in the blood.⁴⁵ The field of targeted nanoparticle therapy is emergent and at present applications are highly limited. This work underscores the potential of targeted nanoparticles in wound and skin care and offers a new targeting platform.

For topically applied nanoparticles, non-specific uptake was significantly reduced and is mostly limited to follicular (passive) penetration.⁴⁶ Active penetration of nanoparticles with size >45 nm is an open and controversial topic of discussion. Although the literature reports variable results on porcine and murine skin showing penetration, there are only two publications that report passive permeation of such particles beyond the stratum corneum.^{46–48} Toll et al.⁴⁸ reports penetration of stratum corneum via hair follicle conduits. However, Kohli and Alpar⁴⁷ attributes penetration of stratum corneum to the strong anionic properties of nanoparticles. If the intent is to target specific cells, high-surface charge of nanoparticles is a confounding factor, as it would be promiscuous and be readily taken up by most cells in proximity.

This work reports a simple and scale-up friendly targeted nanoparticle platform for skin and wound care applications. The design of TLN_κ employed 1,2-dioleoyl-3-trimethylammonium-propane/1,2-dioleoyl-3-dimethylammonium-propane (DOTAP/DODAP) combination pH-responsive lipid components to improve endosomal escape.²¹ Compared to our previously published nanoparticle formulation,²¹ we replaced gramicidin with keratinocyte-targeting ligand to promote cell-specific targeting. To minimize interference of clearance by immune cells, surface charge of anti-miR loaded TLN_κ was designed to be close to neutral. Additional pH versus zeta-potential analyses were conducted to assess the pH dependence of surface charge. TLN_κ possess near-zero surface charges that quickly react to pH drop by boosting surface charges to +15 mV for efficient endosomal release. This design does compromise transfection efficiency but maximizes targeting efficiency, which is desirable in a wound microenvironment complicated by inflammation. By virtue of its design properties, at pH 7 TLN_κ is near zero in charge. This poses stability

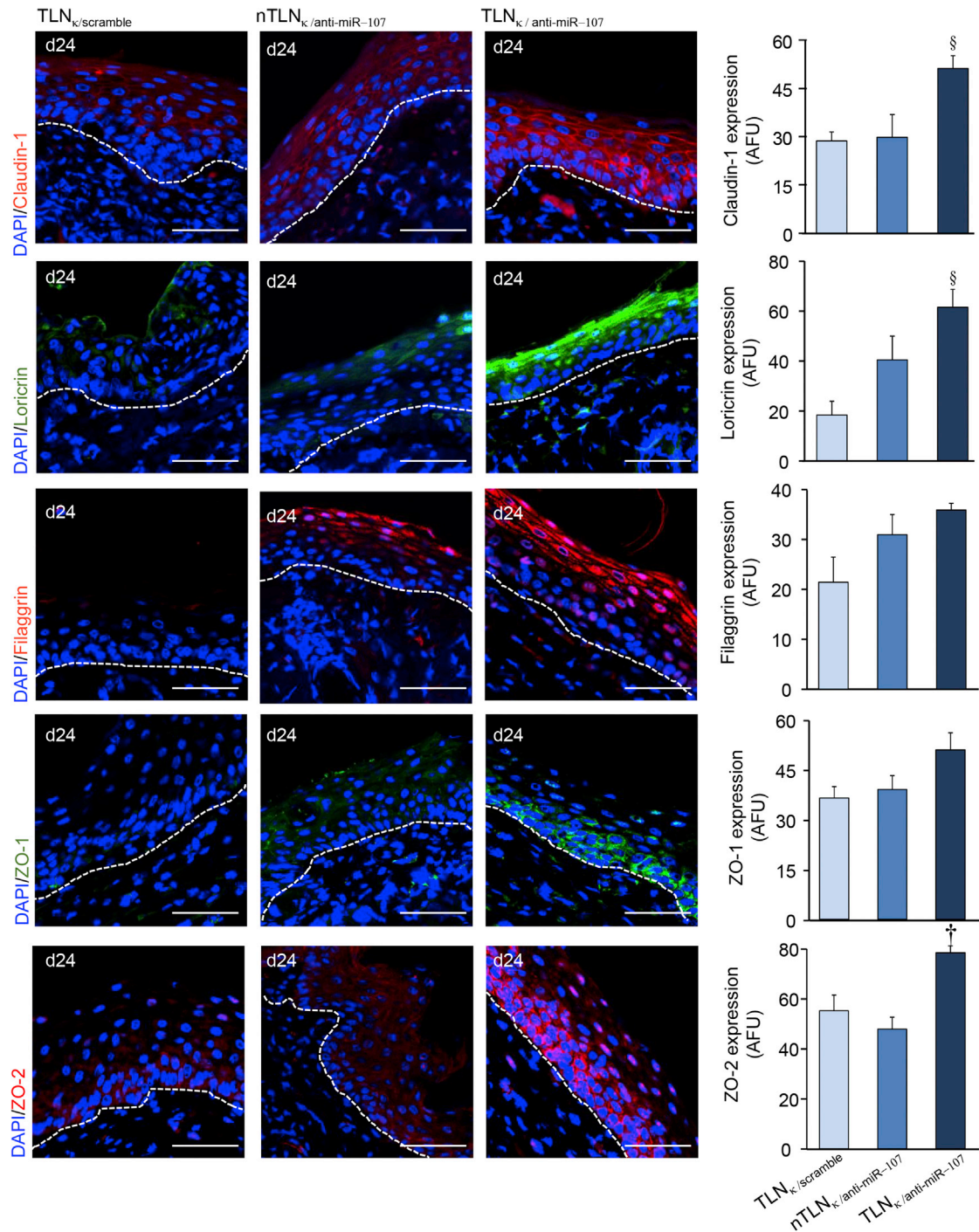


Figure 5. TLN_k/anti-miR-107 Upregulates Epidermal Junctional Proteins

TLN_k/anti-miR-107 increased the expression of claudin-1 (red), loricrin (green), filaggrin (red), ZO-1 (green), and ZO-2 (red) in murine skin at day 24. Sections were counterstained with DAPI (blue). Dermal-epidermal junction is indicated by dashed white line. Scale bars, 50 μm. Abundance of junctional proteins were quantified and expressed graphically as mean ± SEM (n = 4). §p < 0.05, †p < 0.01, ANOVA.

problems due to aggregation, deposition, and sedimentation of the particles.⁴⁹ Lyophilization of TLN_κ not only addresses this limitation but also extends storage life.

Targeted nanoparticles must achieve high uptake efficiency in the desired cell type. The efficiency of uptake is markedly limited in the presence of serum, which may mask targeting ligands by the protein corona.^{43,44,50} In the presence of 10% serum, TLN_κ achieved 90% uptake by keratinocytes within the first 4 hr, demonstrating targeting specificity and efficiency. Rapid uptake by intended cells would minimize clearance by immune cells of the wound microenvironment. In its final form, the lyophilized powder and hydrogel may be stored long-term. A clear translational advantage of TLN_κ is that all material used for its formulation has prior history of FDA approval for human use. Translational development of TLN_κ for human testing is thus warranted.

MATERIALS AND METHODS

1,2-dioleoyl-*sn*-glycero-3-phosphocholine (DOPC), DOTAP, DODAP, and DSPE-PEG₂₀₀₀-Amine were obtained from Avanti Polar Lipids (Alabaster, AL, USA). D- α -Tocopherol polyethylene glycol 1000 succinate (TPGS) and trimethylamine (TEA) were obtained from Sigma-Aldrich (St. Louis, MO, USA). PEGylated bis(sulfosuccinimidyl)suberate (BS(PEG)₅) and Vybrant DiD Cell-Labeling Solution were purchased from Thermo Fisher Scientific (Waltham, MA, USA). A5G33 peptide (sequence, ASKAIQVFLLAG) was custom synthesized by Genscript (Township, NJ, USA). 3M Tegaderm hydrogel was obtained from 3M (St. Paul, MN, USA). Anti-miR-107 and anti-miR-210 with LNA modification as well as negative control A-LNA were synthesized by Exiqon (Woburn, MA, USA).

Lipid-Peptide Conjugation

Lipid peptide conjugation was done by dissolving 4 mg of DSPE-PEG₂₀₀₀-Amine in 500 μ L of DMSO and combined with 5.7 μ L of 250 mM homobifunctional crosslinker (BS(PEG)₅) with addition of 4 μ L of TEA. The reaction proceeded in an oxygen-free environment (nitrogen purge) at room temperature for overnight. Next, 0.5 mg of A5G33 peptide was added into the solution and incubated for 2 hr at room temperature. The molar ratio between DSPE-PEG₂₀₀₀, BS(PEG)₅, and A5G33 peptide was controlled at 1:1.2:1.4. Thin-layer chromatography was used to evaluate the efficiency of conjugation reaction.

Preparation of TLN_κ

TLN_κ were prepared by an ethanol serial dilution method, as described previously.^{51,52} First, lipid components (DOTAP/DODAP/DSPE-PEG₂₀₀₀-A5G33/DOPC/TPGS, 30/24/3/41/2 mol/mol) were dissolved in ethanol. If lipophilic fluorescence dye DiD were chosen to label the LNPs, 0.2% mol/mol amount of dye was added into the above formulation recipe. LNA-based anti-miR was dissolved in equal volume of citric acid buffer (5 mM, pH 4.5) as prepared lipid ethanol solution. The lipid solution was rapidly injected into LNA citric acid buffer at 1:10 anti-miR-

107:total lipids weight ratio. This mixture was further diluted by equivalent volumes (1:1) of HEPES buffer (25 mM, 135 mM NaCl, pH 7.4) twice. Non-targeting LNPs were formulated similarly by substituting DSPE-PEG₂₀₀₀-A5G33 with DSPE-PEG₂₀₀₀-Amine and used as controls. The final product was stored at 4°C.

Lyophilization of TLN_κ

A primary-secondary drying program was employed.⁵³ 1 mL of LNP samples were transferred into 5 mL lyophilization vials (Wheaton, Millville, USA) and diluted with equal volume of 20% sucrose solutions. Slow-freezing processes were chosen: sample vials were partially capped and cooled in a sequence of 2 hr each at 4°C, -20°C, and -80°C. Frozen samples were primary dried at -30°C and 0.12 mbar for 24 hr followed with 25°C and 0.08 mbar secondary drying for an additional 6 hr in a shelf dryer from Labconco (Kansas City, MO, USA). Lyophilized sample stoppers were seated in vacuum and stored at 4°C.

Measurement of TLN_κ Size and Zeta Potential

For mean particle diameter and surface charge measurement, aliquotes of TLN_κ were diluted in PBS (20 mM, pH 7.4). The particle size was determined by dynamic light scattering method using a Zetasizer Nano ZS90 (Malvern, Worcestershire, UK). TLN_κ were dispersed in double-distilled water and tested in volume-weighted size distribution mode. TLN morphology and size distribution were further analyzed by NanoSight NS300 (Malvern, Worcestershire, UK). Aliquots of TLN_κ containing anti-miR-107 were diluted in PBS with a series of pHs (50 mM, from 2 to 11) to determine the pH dependency of surface charge. Zeta potential measurement was carried out on a Zetasizer Nano ZS90 (Malvern, Worcestershire, UK).

Encapsulation Efficiency of TLN_κ

For encapsulation efficiency and stability, TLN_{κ/anti-miR-107} were first diluted into 1 mL solution by 20 mM HEPES buffer and injected into Slide-A-Lyzer Dialysis Cassette (Thermo Scientific, Waltham, MA, USA). A 1 L beaker was filled with 20 mM HEPES buffer. Dialysis cassette was then dropped into the beaker and incubated at room temperature for 4 hr. Encapsulation efficiency was then performed by Quant-iT RiboGreen RNA Kit (Invitrogen, Grand Island, NY, USA). TLN_{κ/anti-miR-107} was lysed with Triton X-100 and the fluorescence intensity (FI) was determined using Multi-Mode Microplate Readers (Biotek, Winooski, VT, USA) at 480 nm λ_{ex} and 520 nm λ_{em} . Loading efficiency was calculated with the following formula:

Encapsulation Efficiency =

$$\left(1 - \frac{\text{Fluorescence without Triton X} - 100}{\text{Fluorescence with Triton X} - 100}\right) \times 100\%.$$

Formulation and lyophilized cake stability was evaluated at -20°C, 4°C, and 25°C over a period of 9 days. The particle size was measured by Zetasizer Nano ZS90.

Cells and Cell Culture

Immortalized human keratinocytes (HaCaT) were grown in Dulbecco's low-glucose modified Eagle's medium (Life Technologies) as described previously.⁵⁴ Human dermal microvascular endothelial cells (HMECs) were cultured in MCDB-131 medium supplemented 10 mM L-glutamine and 100 IU/mL of penicillin and 0.1 mg/mL of streptomycin (Invitrogen), as described previously.⁵⁵ THP-1 are human (pro-) monocytic cell lines that can be differentiated into either dendritic cells or tissue macrophages.⁵⁶ THP-1 cells were differentiated to macrophage-like fate using phorbol 12-myristate 13-acetate (PMA; 20 ng/mL, 48 hr) as previously described.⁵⁷ Differentiated human THP-1 cells were cultured in RPMI complete medium as described previously.^{57,58} Human skin fibroblast BJ cells (ATCC CRL-2522) were obtained from ATCC and were cultured in Eagle's Minimum Essential Medium, (catalog no. 30-2003) as per the instruction provided. All cell cultures media were supplemented with 10% fetal bovine serum (FBS) and 1% antibiotic-antimycotic (AA) (Life Technologies) unless stated otherwise. Cells were maintained in a standard culture incubator with humidified air containing 5% CO₂ at 37°C.

Measurement of Targeting Efficiency by Flow Cytometry

The fluorescence of the cells was determined by using an Accuri C6 flow cytometer (Accuri Cytometers, Ann Arbor, MI, USA). Signals from cells labeled with DiD fluorescent dye were collected on channel FL4 (650 ± 20 nm) after excitation with a 488-nm solid-state laser on a gated population of cells. Data were collected from at least 10,000 cells at a flow rate of 250–300 cells/s. A logarithmic scale was used to measure both background and cell fluorescence. Background fluorescence was then subtracted from cell fluorescence, allowing linear comparisons.

Transfection of miRNA Mimic and Inhibitors

HaCaT cells (0.05 × 10⁶ cells/well in 24-well plate) were seeded for 18–24 hr prior to transfection. HaCaT cells were transfected with either miRIDIAN hsa-miR-107 mimic (50 nM) or hsa-miR-107 inhibitor (100 nM) using DharmaFECT 1 transfection reagent (Thermo Scientific Dharmacon RNA Technologies, Lafayette, CO) per the manufacturer's instructions.^{59–61} miRIDIAN miR mimic/inhibitor negative controls (Thermo Scientific Dharmacon RNA Technologies, Lafayette, CO) were used as control. Cells were collected after 48 hr of miR mimic or 72 hr of miR inhibitor transfection for quantification of miRNA or protein expression. In some experiments, the cells were further reseeded in 96-wells plate or 2-well inserts for cell proliferation or cell migration assay.

Determination of Cell Proliferation

Cell proliferation was measured using a Vybrant MTT Cell Proliferation Assay Kit (Thermo Fisher Scientific) per the manufacturer's instructions as described previously.⁶² At 24 hr after treatment, cells were incubated in fresh low-glucose DMEM culture medium containing 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) for 4–6 h at 37°C with 5% CO₂. After MTT treatment, medium was removed, and DMSO was added (10–20 min at 37°C

with 5% CO₂) to solubilize formazan produced as a result of MTT metabolism. DMSO extract from each well (100 µL) was collected in a 96-well plate, and formazan content was determined by reading absorbance at 540 nm.

Cell Migration Assay

Cell migration assay was performed using culture inserts (IBIDI, Verona, WI) according to the manufacturer's instructions as discussed previously.⁶³ In brief, a confluent cellular monolayer was made in the presence of the insert inside a 24-well plate. Removal of the insert generated a gap in the monolayer. Migration of cells across that gap was studied at different time points following withdrawal of the insert. Images were captured using an inverted microscope (Axiovert 200M, Zeiss, Germany) and analyzed using Zen software.

Animal Model

To generate reproducible, full-thickness burn wounds, an electrically heated burn device with controlled pressure delivery was manufactured. A 0.8-cm diameter stainless steel burning stylus was brought to temperature by an electrically heated element (150 W soldering iron, Great American) inserted into the middle of the stylus. Temperature was controlled by an electronic thermostat (eTRON M, JUMO Process Control, NY), which measured the burn stylus temperature using a thermocouple inserted into the center of the stylus and controlled the amount of current sent to the heating element. Applied pressure was regulated through a compression system comprised of upper and lower aluminum guides connected by spring-loaded guide posts. The heating element can be brought into contact with the murine skin and pressure applied. When maximum pressure is reached, the springs collapse fully and prevent further depression of the burner stylus into the skin below.

Male C57BL/6 mice were obtained from Harlan Laboratory (Indianapolis, IN). Experimental thermal injury was caused by 110°C heated stylus for 15 s. Animals were randomly divided into groups based on a computer generated algorithm (<https://www.random.org/>). Topical application of the LNPs in hydrogel was performed every third day post-burn. All animal studies were performed in accordance with protocols approved by the Laboratory Animal Care and Use Committee of The Ohio State University. During the wounding procedure, mice were anesthetized by low-dose isoflurane inhalation as per standard recommendation. Each wound was digitally photographed at the time point indicated. Wound area was calculated using ImageJ software (NIH, Bethesda, MD).

The animals were euthanized at the indicated post-wounding time point, and wound-edge tissues (2 mm away from the wound) were snap frozen or collected in either optimal cutting temperature compound (OCT) or fixed in formalin for immunohistology.

TEWL

TEWL serves as a reliable index to evaluate the skin barrier function *in vivo*.^{31,64} TEWL was measured from the wounds using Dermalab

TEWL Probe (cyberDERM, Broomall, PA).³¹ The data were expressed in $\text{gm}^{-2} \text{h}^{-1}$.

Laser Capture Microdissection of Epidermis

Epidermal laser capture microdissection (LCM) was performed using the laser microdissection system from PALM Technologies (Bernreid, Germany) as described previously by our group.^{22,65} In brief, sections were stained with hematoxylin for 30 s, subsequently washed with DEPC- H_2O , and dehydrated in ethanol as described previously.⁶⁶ Epidermal fractions were identified based on morphology, cut, and captured under a 20 \times ocular lens. The samples were catapulted into 25 μL of cell direct lysis extraction buffer (Invitrogen). Approximately 1,000,000 μm^2 of tissue area was captured into each cap, and the lysate was then stored at -80°C for further processing.^{22,65}

RNA Extraction and Real-Time qPCR

RNA from mouse wound-edge tissue or cells was isolated using miRVana miRNA Isolation Kit according to the manufacturer's protocol (Ambion Life Technologies) as described previously.⁶⁷ The RNA quality was assessed using Agilent 2100 Bioanalyzer (Agilent Technologies, Santa Clara, CA). Quantification of mRNA expression was done by real-time qPCR using SYBR Green-I.^{65,68} For determination of miR expression, specific TaqMan assays for miRs and the TaqMan miRNA reverse transcription kit were used, followed by real-time PCR using the Universal PCR Master Mix (Applied Biosystems, Foster City, CA).⁵⁸

Histology, Immunohistochemistry, and Microscopy

Histology of skin was performed from 8- μm -thick paraffin sections after staining with H&E. Immunostaining of Dicer (Abcam, ab13502; 1:200), p21^{Waf1/Cip1} (Santa Cruz, sc-397; 1:200), Keratin 14 (Covance, PRB-155P; 1:400), Claudin-1 (Invitrogen, RB9209P; 1:200), loricrin (Covance, PRB-145P; 1:400), filaggrin (Covance, PRB-417P; 1:500), ZO-1 (Invitrogen, 617,300; 1:200), ZO-2 (Invitrogen, 38-9,100; 1:200) were performed on paraffin and cryosections of skin sample using specific antibodies as indicated.⁶⁹ Specificity of the antibodies was validated using rabbit isotype control (Abcam, ab27478; 1:400) (Figure S4). In brief, OCT-embedded tissue was cryosectioned (10 μm), fixed with cold acetone, blocked with 10% normal goat serum, and incubated with specific antibodies overnight at 4°C . Paraffin-embedded tissue were sectioned (8 μm) and deparaffinized, and antigen retrieval was performed prior to blocking. Signal was visualized by subsequent incubation with fluorescence-tagged appropriate secondary antibodies (Alexa 488-tagged α -rabbit, 1:200; Alexa 568-tagged α -rabbit, 1:200).

Data Collection and Statistical Analyses

Samples were coded and data analysis was performed in a blinded fashion. For animal studies, data are reported as mean \pm SD of at least 4–8 animals as indicated. *In vitro* data are reported as mean \pm SD of 3–6 experiments, as indicated in respective figure legends. Student's *t* test (two-tailed) was used to determine significant differences. Comparisons among multiple groups were tested using ANOVA. $p < 0.05$ was considered statistically significant.

SUPPLEMENTAL INFORMATION

Supplemental Information includes four figures and can be found with this article online at <https://doi.org/10.1016/j.yymthe.2018.04.021>.

AUTHOR CONTRIBUTIONS

Conceptualization, J.L., S.G., R.J.L., C.K.S.; Methodology, J.L., S.G., Y.L., R.J.L., C.K.S.; Investigation and Validation, J.L., S.G., M.S.E.M., A.D., Y.L.; Formal Analysis, J.L., S.G., M.S.E.M., S.R.; Writing – Original Draft, J.L., S.G., M.S.E.M., S.R., R.J.L., C.K.S.; Writing – Review and Editing, J.L., S.G., M.S.E.M., A.D., S.R., R.J.L., C.K.S.; Visualization, J.L., S.G.; Funding Acquisition, C.K.S.; Resources, R.J.L., S.R., C.K.S.; Supervision, R.J.L., S.R., C.K.S.

CONFLICTS OF INTEREST

The authors declare no conflict of interest.

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REFERENCES

- Ashton, S., Song, Y.H., Nolan, J., Cadogan, E., Murray, J., Odedra, R., Foster, J., Hall, P.A., Low, S., Taylor, P., et al. (2016). Aurora kinase inhibitor nanoparticles target tumors with favorable therapeutic index in vivo. *Sci. Transl. Med.* 8, 325ra17.
- Dritschilo, A., Huang, C.H., Rudin, C.M., Marshall, J., Collins, B., Dul, J.L., Zhang, C., Kumar, D., Gokhale, P.C., Ahmad, A., et al. (2006). Phase I study of liposome-encapsulated c-raf antisense oligodeoxynucleotide infusion in combination with radiation therapy in patients with advanced malignancies. *Clin. Cancer Res.* 12, 1251–1259.
- Elazar, V., Adwan, H., Bäuerle, T., Rohekar, K., Golomb, G., and Berger, M.R. (2010). Sustained delivery and efficacy of polymeric nanoparticles containing osteopontin and bone sialoprotein antisenses in rats with breast cancer bone metastasis. *Int. J. Cancer* 126, 1749–1760.
- Davis, M.E., Zuckerman, J.E., Choi, C.H., Seligson, D., Tolcher, A., Alabi, C.A., Yen, Y., Heidel, J.D., and Ribas, A. (2010). Evidence of RNAi in humans from systemically administered siRNA via targeted nanoparticles. *Nature* 464, 1067–1070.
- Taberero, J., Shapiro, G.I., LoRusso, P.M., Cervantes, A., Schwartz, G.K., Weiss, G.J., Paz-Ares, L., Cho, D.C., Infante, J.R., Alsina, M., et al. (2013). First-in-humans trial of an RNA interference therapeutic targeting VEGF and KSP in cancer patients with liver involvement. *Cancer Discov.* 3, 406–417.
- Schultheis, B., Strumberg, D., Santel, A., Vank, C., Gebhardt, F., Keil, O., Lange, C., Giese, K., Kaufmann, J., Khan, M., and Drevs, J. (2014). First-in-human phase I study of the liposomal RNA interference therapeutic Atu027 in patients with advanced solid tumors. *J. Clin. Oncol.* 32, 4141–4148.
- Jensen, S.A., Day, E.S., Ko, C.H., Hurley, L.A., Luciano, J.P., Kouri, F.M., Merkel, T.J., Luthi, A.J., Patel, P.C., Cutler, J.I., et al. (2013). Spherical nucleic acid nanoparticle conjugates as an RNAi-based therapy for glioblastoma. *Sci. Transl. Med.* 5, 209ra152.
- Islam, M.A., Reesor, E.K., Xu, Y., Zope, H.R., Zetter, B.R., and Shi, J. (2015). Biomaterials for mRNA delivery. *Biomater. Sci.* 3, 1519–1533.

9. Tolcher, A.W., Rodriguez, W.V., Rasco, D.W., Patnaik, A., Papadopoulos, K.P., Amaya, A., Moore, T.D., Gaylor, S.K., Bisgaier, C.L., Souch, M.P., et al. (2014). A phase 1 study of the BCL2-targeted deoxyribonucleic acid inhibitor (DNAi) PNT2258 in patients with advanced solid tumors. *Cancer Chemother. Pharmacol.* *73*, 363–371.
10. Uchechi, O., Ogbonna, J.D.N., and Attama, A.A. (2014). Nanoparticles for dermal and transdermal drug delivery. In *Application of Nanotechnology in Drug Delivery*, ed., A.D. Sezer, ed. (InTech Open), pp. 193–235.
11. Weissleder, R., Kelly, K., Sun, E.Y., Shtatland, T., and Josephson, L. (2005). Cell-specific targeting of nanoparticles by multivalent attachment of small molecules. *Nat. Biotechnol.* *23*, 1418–1423.
12. Högemann, D., Ntziachristos, V., Josephson, L., and Weissleder, R. (2002). High throughput magnetic resonance imaging for evaluating targeted nanoparticle probes. *Bioconj. Chem.* *13*, 116–121.
13. Bobo, D., Robinson, K.J., Islam, J., Thurecht, K.J., and Corrie, S.R. (2016). Nanoparticle-based medicines: a review of FDA-approved materials and clinical trials to date. *Pharm. Res.* *33*, 2373–2387.
14. Prausnitz, M.R., and Langer, R. (2008). Transdermal drug delivery. *Nat. Biotechnol.* *26*, 1261–1268.
15. Prow, T.W., Grice, J.E., Lin, L.L., Faye, R., Butler, M., Becker, W., Wurm, E.M., Yoong, C., Robertson, T.A., Soyer, H.P., and Roberts, M.S. (2011). Nanoparticles and microparticles for skin drug delivery. *Adv. Drug Deliv. Rev.* *63*, 470–491.
16. Lauterbach, A., and Müller-Goymann, C.C. (2015). Applications and limitations of lipid nanoparticles in dermal and transdermal drug delivery via the follicular route. *Eur. J. Pharm. Biopharm.* *97* (Pt A), 152–163.
17. Desai, N. (2012). Challenges in development of nanoparticle-based therapeutics. *AAPS J.* *14*, 282–295.
18. Shi, J., Kantoff, P.W., Wooster, R., and Farokhzad, O.C. (2017). Cancer nanomedicine: progress, challenges and opportunities. *Nat. Rev. Cancer* *17*, 20–37.
19. Miele, E., Spinelli, G.P., Miele, E., Di Fabrizio, E., Ferretti, E., Tomao, S., and Gulino, A. (2012). Nanoparticle-based delivery of small interfering RNA: challenges for cancer therapy. *Int. J. Nanomedicine* *7*, 3637–3657.
20. Masuda, R., Mochizuki, M., Hozumi, K., Takeda, A., Uchinuma, E., Yamashina, S., Nomizu, M., and Kadoya, Y. (2009). A novel cell-adhesive scaffold material for delivering keratinocytes reduces granulation tissue in dermal wounds. *Wound Repair Regen.* *17*, 127–135.
21. Ghatak, S., Li, J., Chan, Y.C., Gnyawali, S.C., Steen, E., Yung, B.C., Khanna, S., Roy, S., Lee, R.J., and Sen, C.K. (2016). AntihypoxamiR functionalized gramicidin lipid nanoparticles rescue against ischemic memory improving cutaneous wound healing. *Nanomedicine (Lond.)* *12*, 1827–1831.
22. Biswas, S., Roy, S., Banerjee, J., Hussain, S.R., Khanna, S., Meenakshisundaram, G., Kuppasamy, P., Friedman, A., and Sen, C.K. (2010). Hypoxia inducible microRNA 210 attenuates keratinocyte proliferation and impairs closure in a murine model of ischemic wounds. *Proc. Natl. Acad. Sci. USA* *107*, 6976–6981.
23. Texier, I., Goutayer, M., Da Silva, A., Guyon, L., Djaker, N., Jossierand, V., Neumann, E., Bibette, J., and Vinet, F. (2009). Cyanine-loaded lipid nanoparticles for improved in vivo fluorescence imaging. *J. Biomed. Opt.* *14*, 054005, 054005–054011.
24. Wang, B., Zuo, Z., Lv, F., Zhao, L., Du, M., and Gao, Y. (2017). MiR-107 inhibits proliferation of lung cancer cells through regulating TP53 regulated inhibitor of apoptosis 1 (TRIAPI1). *Open Life Sci.* *12*, 200–205.
25. Xia, H., Li, Y., and Lv, X. (2016). MicroRNA-107 inhibits tumor growth and metastasis by targeting the BDNF-mediated PI3K/AKT pathway in human non-small lung cancer. *Int. J. Oncol.* *49*, 1325–1333.
26. Li, X.-Y., Luo, Q.-F., Wei, C.-K., Li, D.-F., Li, J., and Fang, L. (2014). MiRNA-107 inhibits proliferation and migration by targeting CDK8 in breast cancer. *Int. J. Clin. Exp. Med.* *7*, 32–40.
27. Piao, L., Zhang, M., Datta, J., Xie, X., Su, T., Li, H., Teknos, T.N., and Pan, Q. (2012). Lipid-based nanoparticle delivery of Pre-miR-107 inhibits the tumorigenicity of head and neck squamous cell carcinoma. *Mol. Ther.* *20*, 1261–1269.
28. Li, Y., Mao, L., Gao, Y., Baral, S., Zhou, Y., and Hu, B. (2015). MicroRNA-107 contributes to post-stroke angiogenesis by targeting Dicer-1. *5*, 13316.
29. Ristori, E., Lopez-Ramirez, M.A., Narayanan, A., Hill-Teran, G., Moro, A., Calvo, C.F., Thomas, J.L., and Nicoli, S. (2015). A dicer-miR-107 interaction regulates biogenesis of specific miRNAs crucial for neurogenesis. *Dev. Cell* *32*, 546–560.
30. Ghatak, S., Chan, Y.C., Khanna, S., Banerjee, J., Weist, J., Roy, S., and Sen, C.K. (2015). Barrier function of the repaired skin is disrupted following arrest of dicer in keratinocytes. *Mol. Ther.* *23*, 1201–1210.
31. Roy, S., Elgharably, H., Sinha, M., Ganesh, K., Chaney, S., Mann, E., Miller, C., Khanna, S., Bergdall, V.K., Powell, H.M., et al. (2014). Mixed-species biofilm compromises wound healing by disrupting epidermal barrier function. *J. Pathol.* *233*, 331–343.
32. Zatsepin, T.S., Kotelevtsev, Y.V., and Koteliensky, V. (2016). Lipid nanoparticles for targeted siRNA delivery—going from bench to bedside. *Int. J. Nanomedicine* *11*, 3077–3086.
33. D’Mello, S.R., Cruz, C.N., Chen, M.L., Kapoor, M., Lee, S.L., and Tyner, K.M. (2017). The evolving landscape of drug products containing nanomaterials in the United States. *Nat. Nanotechnol.* *12*, 523–529.
34. Davis, M.E., Chen, Z.G., and Shin, D.M. (2008). Nanoparticle therapeutics: an emerging treatment modality for cancer. *Nat. Rev. Drug Discov.* *7*, 771–782.
35. Anselmo, A.C., and Mitragotri, S. (2016). Nanoparticles in the clinic. *Bioeng. Transl. Med.* *1*, 10–29.
36. Gatter, K.C., Brown, G., Trowbridge, I.S., Woolston, R.E., and Mason, D.Y. (1983). Transferrin receptors in human tissues: their distribution and possible clinical relevance. *J. Clin. Pathol.* *36*, 539–545.
37. Stevens, P.J., Sekido, M., and Lee, R.J. (2004). A folate receptor-targeted lipid nanoparticle formulation for a lipophilic paclitaxel prodrug. *Pharm. Res.* *21*, 2153–2157.
38. Li, H., Liu, Y., Chen, L., Liu, Q., Qi, S., Cheng, X., Lee, Y.B., Ahn, C.H., Kim, D.J., and Lee, R.J. (2018). Folate receptor-targeted lipid-albumin nanoparticles (F-LAN) for therapeutic delivery of an Akt1 antisense oligonucleotide. *J. Drug Target.* *26*, 466–473.
39. Chen, W., Zou, Y., Meng, F., Cheng, R., Deng, C., Feijen, J., and Zhong, Z. (2014). Glyco-nanoparticles with sheddable saccharide shells: a unique and potent platform for hepatoma-targeting delivery of anticancer drugs. *Biomacromolecules* *15*, 900–907.
40. Lallana, E., Rios de la Rosa, J.M., Tirella, A., Pelliccia, M., Gennari, A., Stratford, I.J., Puri, S., Ashford, M., and Tirelli, N. (2017). Chitosan/hyaluronic acid nanoparticles: rational design revisited for RNA delivery. *Mol. Pharm.* *14*, 2422–2436.
41. Martínez-Avila, O., Hijazi, K., Marradi, M., Clavel, C., Campion, C., Kelly, C., and Penadés, S. (2009). Gold manno-glyconanoparticles: multivalent systems to block HIV-1 gp120 binding to the lectin DC-SIGN. *Chemistry* *15*, 9874–9888.
42. Chavez-Santoscoy, A.V., Roychoudhury, R., Pohl, N.L., Wannemuehler, M.J., Narasimhan, B., and Ramer-Tait, A.E. (2012). Tailoring the immune response by targeting C-type lectin receptors on alveolar macrophages using “pathogen-like” amphiphilic polyamphiphilic nanoparticles. *Biomaterials* *33*, 4762–4772.
43. Salvati, A., Pitek, A.S., Monopoli, M.P., Prapainop, K., Bombelli, F.B., Hristov, D.R., Kelly, P.M., Åberg, C., Mahon, E., and Dawson, K.A. (2013). Transferrin-functionalized nanoparticles lose their targeting capabilities when a biomolecule corona adsorbs on the surface. *Nat. Nanotechnol.* *8*, 137–143.
44. Harrison, C. (2013). Nanotechnology: biological proteins knock nanoparticles off target. *Nat. Rev. Drug Discov.* *12*, 264.
45. Kang, B., Opatz, T., Landfester, K., and Wurm, F.R. (2015). Carbohydrate nanocarriers in biomedical applications: functionalization and construction. *Chem. Soc. Rev.* *44*, 8301–8325.
46. Schneider, M., Stracke, F., Hansen, S., and Schaefer, U.F. (2009). Nanoparticles and their interactions with the dermal barrier. *Dermatoendocrinol* *1*, 197–206.
47. Kohli, A.K., and Alpar, H.O. (2004). Potential use of nanoparticles for transcutaneous vaccine delivery: effect of particle size and charge. *Int. J. Pharm.* *275*, 13–17.
48. Toll, R., Jacobi, U., Richter, H., Lademann, J., Schaefer, H., and Blume-Peytavi, U. (2004). Penetration profile of microspheres in follicular targeting of terminal hair follicles. *J. Invest. Dermatol.* *123*, 168–176.
49. Jiang, J., Oberdörster, G., and Biswas, P. (2009). Characterization of size, surface charge, and agglomeration state of nanoparticle dispersions for toxicological studies. *J. Nanopart. Res.* *11*, 77–89.

50. Mirshafiee, V., Mahmoudi, M., Lou, K., Cheng, J., and Kraft, M.L. (2013). Protein corona significantly reduces active targeting yield. *Chem. Commun. (Camb.)* *49*, 2557–2559.
51. Yang, X., Koh, C.G., Liu, S., Pan, X., Santhanam, R., Yu, B., Peng, Y., Pang, J., Golan, S., Talmon, Y., et al. (2009). Transferrin receptor-targeted lipid nanoparticles for delivery of an antisense oligodeoxynucleotide against Bcl-2. *Mol. Pharm.* *6*, 221–230.
52. Yung, B.C., Li, J., Zhang, M., Cheng, X., Li, H., Yung, E.M., Kang, C., Cosby, L.E., Liu, Y., Teng, L., and Lee, R.J. (2016). Lipid nanoparticles composed of quaternary amine-tertiary amine cationic lipid combination (QTsome) for therapeutic delivery of AntimiR-21 for lung cancer. *Mol. Pharm.* *13*, 653–662.
53. Abdelwahed, W., Degobert, G., Stainmesse, S., and Fessi, H. (2006). Freeze-drying of nanoparticles: formulation, process and storage considerations. *Adv. Drug Deliv. Rev.* *58*, 1688–1713.
54. Sen, C.K., Khanna, S., Babior, B.M., Hunt, T.K., Ellison, E.C., and Roy, S. (2002). Oxidant-induced vascular endothelial growth factor expression in human keratinocytes and cutaneous wound healing. *J. Biol. Chem.* *277*, 33284–33290.
55. Chan, Y.C., Khanna, S., Roy, S., and Sen, C.K. (2011). miR-200b targets Ets-1 and is down-regulated by hypoxia to induce angiogenic response of endothelial cells. *J. Biol. Chem.* *286*, 2047–2056.
56. Bosshart, H., and Heinzelmann, M. (2016). THP-1 cells as a model for human monocytes. *Ann. Transl. Med.* *4*, 438.
57. Ganesh, K., Das, A., Dickerson, R., Khanna, S., Parinandi, N.L., Gordillo, G.M., Sen, C.K., and Roy, S. (2012). Prostaglandin E₂ induces oncostatin M expression in human chronic wound macrophages through Axl receptor tyrosine kinase pathway. *J. Immunol.* *189*, 2563–2573.
58. Das, A., Ganesh, K., Khanna, S., Sen, C.K., and Roy, S. (2014). Engulfment of apoptotic cells by macrophages: a role of microRNA-21 in the resolution of wound inflammation. *J. Immunol.* *192*, 1120–1129.
59. Chan, Y.C., Roy, S., Huang, Y., Khanna, S., and Sen, C.K. (2012). The microRNA miR-199a-5p down-regulation switches on wound angiogenesis by derepressing the v-ets erythroblastosis virus E26 oncogene homolog 1-matrix metalloproteinase-1 pathway. *J. Biol. Chem.* *287*, 41032–41043.
60. Park, H.A., Kubicki, N., Gnyawali, S., Chan, Y.C., Roy, S., Khanna, S., and Sen, C.K. (2011). Natural vitamin E α -tocotrienol protects against ischemic stroke by induction of multidrug resistance-associated protein 1. *Stroke* *42*, 2308–2314.
61. Gordillo, G.M., Biswas, A., Khanna, S., Pan, X., Sinha, M., Roy, S., and Sen, C.K. (2014). Dicer knockdown inhibits endothelial cell tumor growth via microRNA 21a-3p targeting of Nox-4. *J. Biol. Chem.* *289*, 9027–9038.
62. Rink, C., Gnyawali, S., Stewart, R., Teplitsky, S., Harris, H., Roy, S., Sen, C.K., and Khanna, S. (2017). Glutamate oxaloacetate transaminase enables anaplerotic refilling of TCA cycle intermediates in stroke-affected brain. *FASEB J.* *31*, 1709–1718.
63. Banerjee, J., Das Ghatak, P., Roy, S., Khanna, S., Sequin, E.K., Bellman, K., Dickinson, B.C., Suri, P., Subramaniam, V.V., Chang, C.J., and Sen, C.K. (2014). Improvement of human keratinocyte migration by a redox active bioelectric dressing. *PLoS ONE* *9*, e89239.
64. Pinnagoda, J., Tupker, R.A., Agner, T., and Serup, J. (1990). Guidelines for transepidermal water loss (TEWL) measurement. A report from the Standardization Group of the European Society of Contact Dermatitis. *Contact Dermat.* *22*, 164–178.
65. Roy, S., Patel, D., Khanna, S., Gordillo, G.M., Biswas, S., Friedman, A., and Sen, C.K. (2007). Transcriptome-wide analysis of blood vessels laser captured from human skin and chronic wound-edge tissue. *Proc. Natl. Acad. Sci. USA* *104*, 14472–14477.
66. Kuhn, D.E., Roy, S., Radtke, J., Khanna, S., and Sen, C.K. (2007). Laser microdissection and capture of pure cardiomyocytes and fibroblasts from infarcted heart regions: perceived hyperoxia induces p21 in peri-infarct myocytes. *Am. J. Physiol. Heart Circ. Physiol.* *292*, H1245–H1253.
67. Das, A., Ghatak, S., Sinha, M., Chaffee, S., Ahmed, N.S., Parinandi, N.L., Wohleb, E.S., Sheridan, J.F., Sen, C.K., and Roy, S. (2016). Correction of MFG-E8 resolves inflammation and promotes cutaneous wound healing in diabetes. *J. Immunol.* *196*, 5089–5100.
68. Roy, S., Khanna, S., Rink, C., Biswas, S., and Sen, C.K. (2008). Characterization of the acute temporal changes in excisional murine cutaneous wound inflammation by screening of the wound-edge transcriptome. *Physiol. Genomics* *34*, 162–184.
69. Ahmed, N.S., Ghatak, S., El Masry, M.S., Gnyawali, S.C., Roy, S., Amer, M., Everts, H., Sen, C.K., and Khanna, S. (2017). Epidermal E-cadherin dependent β -catenin pathway is phytochemical inducible and accelerates anagen hair cycling. *Mol. Ther.* *25*, 2502–2512.